

Original Communication

Comparative analysis of the human and zebrafish kinomes: focus on the development of kinase inhibitors

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ABSTRACT

Targeting kinases with semi-selective kinase inhibitors is one of the most successful drug development strategies of the 21st century. Zebrafish have become an increasingly useful model for pharmaceutical development. Watersoluble compounds can be screened for zebrafish phenotypes in a high throughput format against a living vertebrate, and cell-signaling events can be imaged in transparent living fish. Despite zebrafish being a more relevant model than more distantly related systems such as the well-annotated kinome of yeast and drosophila, there is no comparative analysis of the human and zebrafish kinome. Furthermore most approved kinase inhibitors, often called 'DFG in' ATP competitive inhibitors, act on conserved active site residues in the kinase. Since the active site residues can be identified by examining the primary sequence, primary sequence identity can be a rough guide as to whether a particular inhibitor will have activity against another kinase. There is a need to evaluate the utility of zebrafish as a drug development model for active site inhibitors of kinases. Here we offer a systematic comparison of the catalytic domains of classical human kinases with the catalytic domains of all annotated zebrafish kinases. We found a high degree of identity between the catalytic domains of most human kinases and their zebrafish homologs, and we ranked 504 human kinase catalytic domains by order of similarity. We found only 23 human kinases with no easily recognizable homologous zebrafish catalytic domain. On the other hand we found 78 zebrafish kinase catalytic domains with no close human counterpart. These 'additional kinase active sites' could represent potential mediators of zebrafish toxicity that may not be relevant to human kinase inhibitors. We used two clinically approved human kinase inhibitors, one targeting a highly homologous target and one targeting a lesser homologous target, and we compared the known human kinase target structures with modeled zebrafish target structures. As expected, the homologous target had high structural identity, but even the less homologous target had high structural identity in residues contacted by the inhibitor. Overall this analysis should help guide researchers interested in studying human kinases and their inhibitors in more tractable systems.

KEYWORDS: kinome, drug development, kinase inhibitors, human kinases, zebrafish

INTRODUCTION

Protein phosphorylation is the primary signal transduction mechanism in eukaryotic cells. This is accomplished through the opposing action of kinases and phosphatases, and the proper regulation of these enzymes is critical for cell homeostasis [1-3]. Inhibiting kinases has become

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one of the most reliable pharmacologic strategies in the last decade with several new kinase targeting drugs currently in use [4]. Initially kinase inhibitors were intensively studied and approved as cancer drugs, but now the scope of their potential use has widened to immune regulation, anti-infectives, and cardiovascular diseases [4, 5]. Tofacitinib is currently approved for rheumatoid arthritis, and other Jak family inhibitors are entering clinical trials for rheumatoid arthritis and psoriasis [6]. Inhibitors for p38 MAP kinases are under investigation for treatment of atherosclerosis and chronic obstructive pulmonary disease (COPD) [7]. These and other successes have reinforced the search for small molecule kinase inhibitors which could selectively inhibit kinases with minimal side effects. There are 518 human kinases and inhibition of any of these kinases can potentially elicit a potent therapeutic effect because of the intrinsic cascading mechanism each kinase possesses [5]. Initially there was concern and skepticism that small molecules could confer selectivity for a single kinase. It is now clear that sufficient selectivity is achievable [5, 8]. Therapeutics inhibiting more than one kinase are sometimes desirable, and potentially more than 518 different effects could be achieved by inhibiting different combinations of kinases. Currently 25 kinase inhibitors are FDA approved, most of which are oral [4].

While the so-called 'atypical' kinases lack canonical primary sequence motifs, there is a common evolutionary and structural basis for protein kinases [9]. Protein kinases have a bilobed structure with catalysis occurring in the cleft between lobes [10]. The cleft contains key conserved features critical to phosphorylation: an ATP binding pocket, substrate binding residues, and a divalent cation-typically Mg²⁺ [10, 11]. Throughout the kinome there are well-recognized conserved motifs that collectively line the cleft, coordinate the metal cofactor and transfer the phosphate from ATP to the substrate [10, 11]. Steven Hanks and others identified several domains which were conserved among almost all kinases and were essential for their kinase activity [10, 12]. These domains became subsequently known as Hanks domains (Figure 1). As the ancestral kinases diversified, many different subfamilies evolved specializing in, for example,

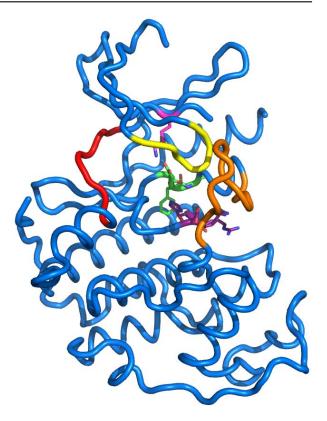


Figure 1. CDK2 (4EK3) as an example of a classical Hank's kinase with critical Hanks motifs highlighted. Hinge region in red, P-loop in yellow, activation loop in orange, invariant lysine in magenta sticks, HRD in purple sticks, and DFG in green sticks.

tyrosine phosphorylation or serine/threonine phosphorylation [13]. The overall human kinome has been divided into subfamilies based on structural and functional differences, and this classifying information is annotated in online databases [1, 14]. Prominent families include serine/ threonine kinases such as AGC and CAMK as well as exclusive tyrosine kinases (TKs) and tyrosine kinase like (TKL) kinases which phosphorylate tyrosine as well as serine and threonine [15]. How these kinase families evolved has been examined in model organisms across a vast swath of divergent organisms from Saccharomyces to mice to humans, but more recent model organisms such as zebrafish have not been analyzed in detail [16].

Despite extensive diversification, the Hanks domains have remained largely unchanged, particularly inside the catalytic domain of the N-lobe between the glycine rich 'P-loop' and the DFG motif [10]. In fact, even small changes in these motifs lead to the categorization of kinases as likely pseudokinases [17]. Some classified pseudokinases retain catalytic activity, but non-canonical motifs at least suggest functions other than phospho-transfer [17, 18]. Although the Hanks domains in the cleft are highly conserved, some variation near the motifs may be present among species, particularly if their substrates diverged greatly. To better utilize zebrafish as a high throughput model for kinase inhibitor drug development, it is necessary to evaluate how similar the clefts are for both the human and zebrafish kinome.

Zebrafish (Danio rerio) is now a well-established model organism for developmental biologists [19]. It has many advantages including low cost, tissue transparency, fecundity, and short generation time [20]. These advantages have also made zebrafish an attractive model for drug development [21]. The zebrafish genome is sequenced and it has 71% homology to the human genome [22]. Genome duplication events in the teleost lineage complicate genetic comparisons between zebrafish and humans, since some genes were duplicated several times and others were not at all [23]. A common tool to probe the specific genetics of zebrafish has been the use of antisense morpholino oligomers to knock down specific genes; however, the specificity of this has been challenged recently [24]. The use of small molecule inhibitors may be a complementary approach if key inhibitor binding sites are well conserved between humans and zebrafish. The zebrafish kinome has yet to be extensively described, but if the kinome parallels the genome in homology, then the zebrafish may potentially serve as a functional model organism for kinase inhibitor development. While zebrafish are already a model for testing drug toxicity [21], expanding zebrafish utility as a kinase-specific model for toxicity and efficacy will require more detailed knowledge about the zebrafish kinome.

Understanding the relatedness of human and zebrafish kinases will improve the reliability of zebrafish as a model organism for studying kinase inhibitors. Testing drugs for human kinases with no clear zebrafish homolog might yield false negatives, and conversely, zebrafish kinases which have clear homologs to human kinases but with several specialized isoforms not found in humans may also provide data that is ultimately not helpful in the drug development process. Knowledge of these relationships would streamline drug development by identifying candidate kinase targets which are viable models in zebrafish and allowing more high-throughput testing to be done earlier in the drug development process. Here we did not examine the overall kinase homology *per se* but specifically analyzed how well the active sites match since most ATP competitive kinase inhibitors rely on critical residues in the Hank's domains.

METHODS

The human kinase domain sequences were retrieved from the KinBase database [1]. There are 531 non-redundant human kinases annotated in Kinbase, of which 25 are classified as atypical. Manning and colleagues have identified 15 additional atypical kinases and 13 pseudokinase domains, of which the former were excluded, and the latter were included in our analysis [1]. Approximately 350 zebrafish kinases were obtained from ensemble [25] and approximately 550 were obtained from zfin [26] using pfam 00069 for a total of ~900 zebrafish sequences from both sources. These kinases were visually examined, and redundant sequences were removed to arrive at a final list of 692. The shortest canonical human kinases from each family were used as a model to identify zebrafish kinase catalytic domains by performing sequence alignment and trimming zebrafish sequences where they align to the human catalytic domains. MultAlign [27] was used to align the kinase domains of the organisms up within their respective subfamilies in order to identify the homologous kinase domains in zebrafish kinases, and analysis was restricted to the kinase domain. Residues prior to the start of the P-loop, usually indicated by the presence of a nearby glycine rich region, and residues subsequent to the DFG motif were truncated, based on the alignment to the canonical kinase domain. This region will be subsequently referred to as the 'catalytic domain' (CD) (Figure 1).

Clustal Omega [28] was used to perform a protein BLAST analysis and generate a phylogenetic tree that depicted the relationship between the human and zebrafish kinase CDs. The data were organized by pairing kinases grouped between species and unique CDs that have no orthologs between humans and zebrafish. The maximum matched percent identity of each zebrafish CD and of each human CD was extracted using Excel (Microsoft) and condensed into a master table (Table S1). A frequency distribution of the maximum percent identities was calculated using Prism (GraphPad software). A cutoff of 40% identity was chosen based on the frequency distribution of all maximum CD percent identities (Figure S1) as well as based on visual examination of the dendogram. Below 40% identity there was not clear pairing of human and zebrafish kinase active sites.

Crystal structures of FDA approved kinase inhibitors were downloaded from the PDB (3OG7 and 3LXK) [29, 30] and visualized in Pymol (Schrodinger). Predicted zebrafish kinase models were generated using I-Tasser [31]. Structural figures were also rendered in Pymol.

RESULTS AND DISCUSSION

The zebrafish and human kinase catalytic domains share high identity

The protein BLAST analysis revealed generally high percent identities between the zebrafish and

human kinase catalytic domains (CD). Four hundred and eighty one human CDs matched to 614 zebrafish CDs with a percent identity greater than 40 (Figure 2). The extra zebrafish kinases are primarily due to additional isoforms for highly homologous forms of certain kinases—chiefly the PIM kinases and Aurora kinases (Table S1). Twenty four unique or divergent human CDs were identified (Figure 2, Table 1) and 78 unique zebrafish CDs were identified (Figure 2, Table 2). The average percent identify of CDs between both kinomes including unique kinases is 69% with a median identity of 75%, suggesting the identity of the kinome corresponds well to the total genomic identity [22].

Unique zebrafish CDs consist of many alternate isoforms of homologous kinases

There are 78 unique zebrafish CDs, defined as those with no human CD of greater than 40% identity (Table 2). Several of these CDs match as isoforms of zebrafish CDs which have percent identities greater than 40% (notably the PIM family and Aurora family); however, some of these CDs are isoforms of similar CDs with no strong human protein homolog (such as the TSSK family). This 'soform variation' may be a function

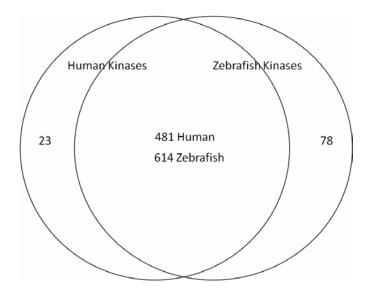


Figure 2. Unique vs. shared kinase catalytic domains. Of the total 692 non-redundant zebrafish sequences analyzed, 614 had greater than 40% identity to 481 human kinase catalytic domain sequences. Only 23 human sequences were less than 40% identity to any zebrafish sequence and 78 zebrafish sequences were less than 40% identity to any human sequence.

Table 1. Divergent or unique human kinase catalytic domains. Human catalytic domain sequences which do not match any zebrafish sequence with greater than 40% identity are shown below. Several of the unique sequences are members of distinct kinase families.

Unique human kinases	Percent identity	Zebrafish kinase match
СОТ	39	CHUK
FAM20A	24	MINK1
FAM20B	18	OXSR1B
FAM20C	26	PIM1
FJB1	27	PINK1
GCN2-P	31	MARK1
MLKL	37	TAOK1A
NME1A	21	SIK2A
NME1B	21	JAK1
NME3	20	PRKD3
NME4	21	JAK1
NME5	21	GUCY2F
NME6	21	РОМК
NME7a	20	ZGC66101
PAN3	26	DCKL1B
SgK223	20	PRKD3
SgK269	25	PRKD3
SgK396	33	MAP3K12
SGK495	30	PTK6B
Slob	31	CDK13
TXNDC3	23	MYLKA
TXNDC6	19	CAMKVL
ULK4	39	STK36

of the diverged habitat of human and zebrafish (land vs. water) or some may be remnants of genome duplication events. These kinases are categorized as isoforms in this study based solely on maximum percent identity; therefore further functional characterization of these kinases is needed if they are to be investigated as drug target models.

Only 5% of human CDs are unique

The unique human CDs form a group of only 24 (Table 1). Interestingly, several of these CDs are related to each other, such as the FAM family, the NME family, and the SgK family. The FAM20 kinases are involved in phosphorylation of secreted substrates [32]. Given that humans and zebrafish live in fundamentally different environments, it is not surprising that pathways which interact with the environment (i.e. secretion) would be divergent. The NME family of kinases has been implicated in the function of tumor metastasis; however, the function of these kinases has been scarcely studied outside of humans [33]. Some pseudokinases such as sgk495 are represented, but unlike the FAM20 and NME kinase families, the SGK subfamily has several members with homologs between human and zebrafish (Table S1). The SGKs are generally involved in stress responses and cellular channel activation [34]. It is possible that some of these channel signaling processes have evolved to be highly divergent in humans, but several of these kinases are still under-explored.

There are seven typical human kinases that have no ortholog in mice [35]. Interestingly, while for many of these 7 there is likely not a strict zebrafish homolog, there is a CD with high identity to the human kinase 'missing' from the mouse kinome. For example neither the mouse kinome nor the zebrafish kinome has a CDK3 ortholog, but the CD of human CDK3 is 80% identical to that of zebrafish CDK2 (Table S1). Another example is that for human CK1alpha2: no ortholog is present in mice, but a high identity (98%) CD is present in zebrafish. The weakest zebrafish match among the 7 human CDs not present in mice is for human PKSH2 at only 74%. The differences in mouse and zebrafish kinomes suggest that organism selection for a particular drug-target model is critically dependent on information gleaned by the type of analysis presented in this study.

Pseudokinases are generally less conserved between zebrafish and humans

Forty-eight human kinases have been classified as 'pseudokinases' on the basis of variation in one of the three Hank's domains (VAIK, HRD, or DFG) necessary for catalysis, and 28 have conserved homologues in yeast, worms, mice, and humans [17]. **Table 2.** Catalytic domains of divergent or unique zebrafish kinases. Zebrafish catalytic domain sequences which do not match any human sequence with greater than 40% identity are shown below. Most (72/78) unmatched kinases are mutants or splice variants of otherwise well matched partners. Six of the catalytic domains are likely truly unique.

Unique zebrafish kinases	Percent identity	Human kinase match
SICH211-138G9.2	39	AurB
SICH211-138G9.3	39	AurB
SICH211-272B8.7	38	AurB
SIDKEY-22I16.10	39	AurB
SIDKEY-34D22.3	38	AurB
SIDKEY-80C24.1	39	AurB
SIDKEY-80C24.5	39	AurB
SIDKEYP-67E1.2	39	AurB
SIDKEY-183G16.2	38	HUNK
SIDKEY-206F10.6	39	MARK1
SIDKEY-83M22.16	38	MARK1
PAK7	26	MLK1
SIDKEY-155D18.1	37	MSK2A
SIDKEYP-67E1.4	39	MSK2A
РХК	24	NuAK1
SIDKEY-197D18.1	39	NuAK1
SIDKEY-197D18.2	39	NuAK1
G6PD	19	PAK4
SICH211-249H16.8	38	PASK
SICH211-57M13.8	38	PASK
SIDKEY-211E20.1	38	PASK
SIDKEY-37M8.14	39	PASK
SICH211-119D14.4	38	PIM1
SICH211-168M18.3	33	PIM1
SICH211-196G2.7	34	PIM1
SICH211-196G2.8	38	PIM1
SICH211-198E20.11	38	PIM1
SIDKEY-236A14.6	38	PIM1
SIDKEYP-100A1.5	37	PIM1

Table 2 continued..

Unique zebrafish kinases	Percent identity	Human kinase match
SICH211-176G13.9	39	PIM2
SICH211-215M21.11	39	PIM2
SICH211-237D5.4	39	PIM2
SICH211-57M13.10	36	PIM2
SIDKEYP-110E4.3	39	PIM2
SICH211-147H1.4	39	PIM3
SICH211-196N4.5	32	PIM3
SICH211-196N4.7	35	PIM3
SICH211-196N4.8	32	PIM3
SICH211-214C11.2	35	PIM3
SICH211-214C11.4	32	PIM3
SICH211-214C11.7	31	PIM3
SICH211-214C11.8	32	PIM3
SICH211-57M13.3	39	PIM3
SICH211-57M13.5	39	PIM3
SICH211-57M13.6	39	PIM3
SICH211-57M13.7	39	PIM3
SIDKEY-222B8.5	34	PIM3
SIDKEYP-67E1.6	39	PIM3
SIDKEY-211E20.8	39	PKD1
SIDKEY-211E20.9	39	PKD1
SICH211-57M13.9	36	SIK
SICH211-10J20.2	38	TSSK1
SICH211-10J20.4	37	TSSK1
SICH211-215M21.13	37	TSSK1
SIDKEY-206F10.7	39	TSSK1
SIDKEY-217M5.9	38	TSSK1
SIDKEY-197D18.3	39	TSSK2
SICH211-160D20.5	38	TSSK3
SICH211-202N12.2	37	TSSK3
SICH211-207C6.4	37	TSSK3
SICH211-238G23.1	38	TSSK3
SICH211-57M13.1	36	TSSK3

Unique zebrafish kinases	Percent identity	Human kinase match
SIDKEY-248F6.3	39	TSSK3
SIZFOS-754C12.2	38	TSSK3
SICH211-12E13.5	38	TSSK4
SICH211-12E13.6	38	TSSK4
PRKD3	27	TTBK2
PKN1B	23	TXNDC3

Table 2	continued
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Although these 48 have atypical variation in key residues based on primary sequence, several have been shown to be catalytically active [17, 18]. For example, WNK1 does not have the classical VIAK motif in β -strand 3, but instead has a lysine residue (K233) that enters the active site from β -strand 2 and confers catalytic activity [36]. This suggests that structural and functional analysis is key to pseudokinase classification [18]. The CDs of twenty-two of these pseudokinases are also conserved between zebrafish and humans (Table 3). Of these 22, differences between zebrafish and human pseudokinases owing to the loss of critical Hanks domains (DFG, HRD, and VIAK) were only seen in three: Trib1, Trib2, and Sgk494A (Table 3). The zebrafish Trib kinases deviate more from the classical Hank's domains than do their human equivalents; however, Sgk494A has retained all of these domains. This suggests that there may be potential differences in structure and function between some of these pseudokinases which evolved after the human/zebrafish divergence. In addition to unique human pseudokinases, several groups which are represented in zebrafish have additional isoforms in humans (e.g. ANPa/b and RSKL1/2) further suggesting additional roles in humans that may be accomplished by fewer isoforms in zebrafish or not at all.

Structural identity is conserved in active sites of varying sequence identity

Although considerable attention is given to sequence identity in the literature, structural identity may be a more accurate predictor of functional conservation [8]. Fortunately, several of the FDA approved kinase inhibitors have been crystallized with their targets, and structures are available [4, 8]. The identity of the targets of these FDA approved drugs is variable, but analysis of these structures can provide information as to whether the most critical of contacts is preserved.

The FDA approved kinase inhibitor vemurafinib is a BRAF kinase inhibitor [4, 29]. Human BRAF kinase has 99% identity to zebrafish BRAF kinase in the catalytic active site (Table S1). Vemurafinib is coordinated in the BRAF active site by hydrogen bonding to the hinge region backbone and the activation loop backbone as well as through several stacking interactions and hydrophobic interactions (Figure 3). The BRAF kinase CDs are essentially identical between human and zebrafish; therefore the drug is almost certain to bind to zebrafish BRAF as it would in human BRAF.

Tofacitinib was FDA approved for arthritis and it inhibits human Jak3 kinase [4, 30]. Jak3 has one canonical CD and one pseudokinase domain. The canonical kinase domain has a 68% identity to zebrafish Jak2A (Table S1). Despite the lower sequence identity in the CDs, they still maintain high structural similarity (Figure 4). Tofacitinib binds to the active site of Jak3's canonical CD similar to most ATP competitors by hydrogen bonding to the hinge region backbone (Figure 4). It is also stabilized by hydrogen bonding to the P-loop and through hydrophobic interactions at the base and back of the active site. Although the CD residues appeared unchanged from the primary sequence, the higher overall divergence may have created conformational changes not evident from primary sequence alone. The I-Tasser-predicted structure shows the residues pointing toward the catalytic site in the CD and overall conformation are mostly unchanged (Figure 4). There is a small kink introduced in human Jak3 by substituting an alanine for glycine at aa966 (152), and there is a cysteine to leucine conversion at aa909 (142). Neither of these changes appears to affect the coordination of the drug in the active site, suggesting that despite a larger amount of evolutionary divergence compared to BRAF, the Jak3 active sites are structurally conserved and would be accurate models for testing new drugs. The structures of many human kinases are known, and several have been co-crystallized with drugs [8]. The sequence analysis presented here, combined **Table 3.** Pseudokinase domain divergence. Of the 48 known human pseudokinase catalytic domains, 22 have various degrees of identity to zebrafish proteins. Pseudokinases with no clear homologous non-redundant partner are left unmatched. Modified critical Hanks domains (DFG, HRD, or VIAK) are indicated based on Boudeau and colleagues [17] and alignment of the matching pairs.

Human pseudokinase	Modified domains	% identity	Zebrafish pseudokinase	Modified domains
ANPA	HRD	84	NPR1A	HRD
ANPB	HRD			
CASK	DFG	99	CASKA	DFG
CCK4	DFG			
CYGD	HRD	60	GC3	HRD
CYGF	HRD	77	GC2	HRD
EPHA10	DFG, HRD, VIAK			
EPHB6	DFG, HRD, VIAK			
GCN2-P	HRD, VIAK			
HER3 (ERBB3)	HRD	75	ERBB3A	HRD
HSER	HRD			
ILK	DFG, HRD	86	ILK	DFG, HRD
IRAK2	DFG, HRD			
JAK1-P	HRD	70	JAK1-P	HRD
JAK2-P	HRD	83	JAK2B-P	HRD
JAK3-P	HRD	60	JAK3-P	HRD
KSR1	VIAK			
KSR2	VIAK	88	KSR2	VIAK
MLKL	DFG, HRD			
NRBP1	DFG, HRD, VIAK	95	NRBP1	DFG, HRD, VIAK
NRBP2	DFG, HRD, VIAK			
PSKH2	HRD			
RSKL1	DFG, VIAK			
RSKL2	DFG, VIAK			
SCYL1	DFG, HRD, VIAK			
SCYL2	DFG, HRD, VIAK	89	SICH211-244B2.1	DFG, HRD, VIAK
SCYL3	DFG, HRD, VIAK	73	SCYL3	DFG, HRD, VIAK
SGK071	HRD			
SGK196	DFG, HRD, VIAK	60	РОМК	DFG, HRD, VIAK
SGK223	DFG			
SGK269	DFG			

Human pseudokinase	Modified domains	% identity	Zebrafish pseudokinase	Modified domains
SGK307	DFG, HRD			
SGK396	HRD			
SGK494	DFG, HRD, VIAK	59	SGK494A	NONE
SGK495	DFG			
SLOB	DFG, HRD, VIAK			
STLK5 (STRADA)	DFG, HRD, VIAK	72	STRADA	DFG, HRD, VIAK
STKL6 (STRADB)	DFG, HRD			
SURTK106	DFG			
ТВСК	DFG, HRD, VIAK	62	TBCK	DFG, HRD, VIAK
TRB1	DFG	55	TRIB1	HRD, VIAK
TRB2	DFG	58	TRIB3	DFG, HRD, VIAK
TRB3	DFG			
TYK2-P	HRD	64	TYK2-P	HRD
ULK4	DFG, VIAK			
VACAMKL	HRD	91	CAMKVA	HRD
VRK3	DFG, HRD, VIAK	46	VRK3	DFG, HRD, VIAK

Table 3	continued
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with structural information from published and modeled drug-kinase interactions, provide potentially insightful tools for drug development.

Limitations

There are several limitations to both our analysis and the general use of zebrafish as an alternative to more costly mammalian models for kinase inhibitor drug development. Testing drugs in zebrafish requires a certain amount of water solubility. Solvents such as DMSO, used to deliver insoluble drugs have toxicity effects on their own, reducing the effectiveness of a freshwater organism model. Fortunately, most kinase inhibitors are less than 1000 Daltons in size and have some polar moieties, allowing them to have sufficient water solubility [8], and all 25 of the currently FDA approved kinase inhibitors are orally available [4].

The zebrafish genome is rapidly improving in annotation but is certainly not as comprehensively annotated as the human genome. For example, twice the zebrafish kinases from zfin were recovered as were in ensemble, but there could still be unannotated kinases that were not included in this study. Very few human kinases were not matched to a zebrafish kinase in this analysis, which could indicate the coverage is sufficient and additional zebrafish kinases would likely be used for biological processes specific to the zebrafish.

Although this work has presented divergent or unique zebrafish CDs based on a histogram of identity and an alignment, the specific utility of zebrafish for drug development for a specific human kinase depends on a number of factors beyond the cut-off presented here. The zebrafish kinome consists of several isoforms of kinases with moderate-to-high identity to their human These isoforms counterpart. are mostly uncharacterized and may be unique to zebrafish and have unique functions. Attempts at drug development targeting kinases which have several other isoforms may be clouded by off target effects. Careful scrutiny should be made in using

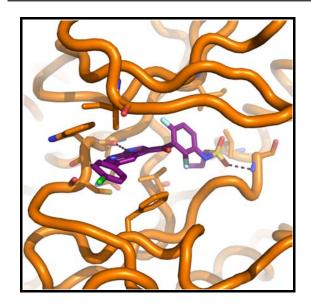


Figure 3. Vemurafenib in human BRAF kinase. The structure of human BRAF with bound Vemurafenib (3OG7) is shown in orange cartoon tube with critical coordinating residues shown in orange sticks. Vemurafenib is shown in purple sticks and hydrogen bonds to the drug are shown as black dashes. Vemurafenib is an FDA approved kinase inhibitor which selectively blocks human BRAF kinase activity and is indicated to treat some melanomas [4, 29]. The zebrafish BRAF kinase catalytic domain has 92% identity to human BRAF. Alignment of the zebrafish sequence to the human structure (3OG7) indicates that all drug coordinating residues in human BRAF are identical to zebrafish BRAF.

zebrafish as a model for a particular kinase for which zebrafish possesses multiple isoforms.

This analysis focused only on primary sequences. Although the structures of the zebrafish kinases are predicted to be similar to their human counterparts based on primary sequences and known similar structures, subtle nuances may still create situations where certain inhibitor interactions are disrupted or strengthened. The quaternary interactions of human kinases have been elucidated thoroughly in some cases such as cyclins and CDKs [37], but this is generally lacking in zebrafish. It is possible that species-specific accessory proteins could cause appreciable conformational changes in the structures of these kinases and alter potential binding patterns. As discussed above, the role of pseudokinases in zebrafish is not well known, and allosteric activation by or of pseudokinases is generally underexplored [17, 18]. Additional

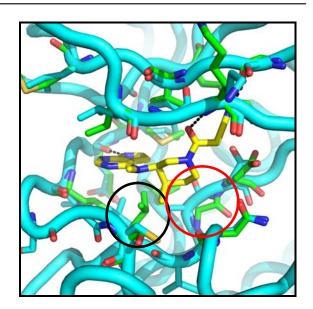


Figure 4. Tofactinib in human JAK3 kinase. The structure of human JAK3 with bound Tofactinib is shown in cyan cartoon tube with critical coordinating residues shown in cyan sticks (3LXK). Tofactinib is shown in yellow sticks. Zebrafish JAK2B residues are shown as green sticks. Tofactinib hydrogen bonds are shown as black dashes. Tofactinib is an FDA approved kinase inhibitor which selectively blocks human JAK3 kinase activity and is indicated to treat rheumatoid arthritis [4, 30]. The zebrafish JAK2A kinase catalytic domain has 68% identity to human JAK3. Threading the zebrafish sequence to the human JAK3 structure (3LXK) indicates that all but two residues in the human catalytic pocket JAK3 are identical to zebrafish JAK2A: C909-Hs is equivalent to L142-Dr (circled in black) and A966-Hs is equivalent to G152 (circled in red).

structural information will be needed in the future to address these concerns.

CONCLUSIONS

Overall, this work should facilitate decisions on when zebrafish represent a useful choice to study human kinase inhibitors. The utility clearly varies from high for inhibitors targeting highly homologous kinase CDs (BRAF) and subtly divergent CDs (Jak3), to low for more highly divergent kinases (FAM20 kinases) or if several divergent isoforms (PIM kinases) exist.

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CONFLICT OF INTEREST STATEMENT

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SUPPLEMENTARY FIGURE AND TABLE

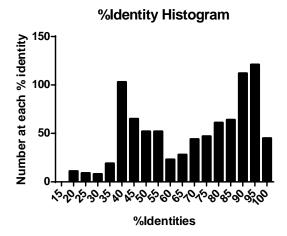


Figure S1. Frequency distribution of all kinase % identities. The percent identity of all human and zebrafish kinase catalytic domains was plotted by frequency. A significant break at 40% identity was observed, suggesting this as a cutoff to delineate unique or divergent catalytic domains.

Table S1. Highest percent identity match of all humankinases with all zebrafish kinases.

Human kinase	% identity	Zebrafish kinase
AAK1	89	AAK1A
	88	AAK1B

Human kinase	% identity	Zebrafish kinase
ABL1	90	ABL2
ABL2	94	ABL2
ACK	62	TNK1
A CEED 2	83	ACVR2AA
ACTR2	82	ACVR2AB
ACTR2B	68	ACVR2B
AKT1	94	AKT1
A 1/700	95	AKT2
AKT2	73	AKT2L
A 1/700	99	AKT3
AKT3	99	AKT3A
ALK	45	TEC
ALK1	81	ACVRL1
	83	ACVR1L
ALK2	82	ACVRL1
	96	ACVR1B
ALK4	96	ACVR1BA
	95	ACVR1BB
ALK7	89	TGFBR1A
AMPKA1	96	PRKAA1
AMPKA2	72	PRKAA2
	89	RIPK4
ANKRD3	72	ANKK1
ANPa	84	NPR1A
ANPb	81	NPR1A
ARAF	86	ARAF
AurA	84	AURKA
	79	AURKB
	40	SICH211-
	40	280A23.2 SICH73-21B16.2
AurB	40	SICH73-21B10.2 SICH73-21B16.3
		SICH75-21B16.5 SIDKEYP-
	39	67E1.2
	39	SIDKEY- 80C24.1

Table S1	continued
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Human kinase	% identity	Zebrafish kinase
	20	SIDKEY-
	39	80C24.5
	20	SIDKEY-
	39	22I16.10
	39	SICH211-
AurB		138G9.2
	39	SICH211-
		138G9.3 SIDKEY-
	38	34D22.3
	29	SICH211-
	38	272B8.7
	77	AURKB
	46	SIDKEY-
	rO	44G23.9
	41	SICH211-
		122L14.3 SICH211-
	41	122L14.7
	4.1	SICH73-
AurC	41	132G21.6
	41	SIRP71-84D9.6
	41	SICH73- 313G15.1
	41	
		SIRP71-84D9.4
	41	SIDKEY-40J3.1
	41	SIRP71-1L17.1
AXL	62	TYRO3
BARK1	98	ADRBK2
BARK2	98	ADRBK2
BIKE	81	BMP2K
BLK	83	BLK
BMPR1A	91	BMPR1AA
	89	BMPR1AB
BMPR1B	94	BMPR1BA
Dim Kib	92	BMPR1BB
BMPR2	88	BMPR2B
Divit K2	71	BMPR2A
BMX	59	TEC
BRAF	99	BRAF
BRK	51	PTK6B

Human kinase	% identity	Zebrafish
	78 Identity	kinase
BRSK1	96	SIDKEY-
DIGITI	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	16P21.7 SIDKEY-
	97	16P21.7
DDGWA	~ ~	SIDKEY-
BRSK2	97	16P21.7
	96	SICH211-
		255P10.4
BTK	65	TEC
BUB1	43	CHEK1
BUBR1	40	PHKG1B
CAMK1A	83	CAMK1A
CAMK1B	67	PNCK
	89	CAMK1DA
CAME 1D	86	CAMK1DB
CAMK1D	86	CAMK1DBA
	83	CAMK1A
CAME 1C	79	CAMK1GB
CAMK1G	74	CAMK1GA
	97	CAMK2A
CAMK2A	95	CAMK2D1
	43	SICH211-
	15	151H10.1
CAMK2B	96	CAMK2B1
C/ IIVIK2D	96	CAMK2B2
CAMK2D	96	CAMK2D2
CAMK2G	96	CAMK2G1
CAMK4	93	CAMK4
C ₂ MKV1	80	CAMKK1
CaMKK1	80	CAMKK1A
	73	CAMKK1B
CaMKK2	72	SICH73-
CANGOV		62L21.1
CAMLCK	77	MYLK3
~	99	CASKA
CASK	99	CASKAB
	98	CASKB
CCK4	41	MUSK

Human kinase	% identity	Zebrafish kinase
CCRK	82	CDK20
CDC2	86	CDK1
CDC7	69	CDC7
CDK10	89	CDK10
CDK11a	98	CDK11B
CDK19	99	CDK19
CDK2	89	CDK2
CDK3	80	CDK2
CDK4	75	CDK4
CDK4	53	CDK21
CDK5	97	CDK5
CDK6	92	CDK6
CDK7	92	CDK7
CDK8	99	CDK8
CDK9	93	CDK9
CDKL1	92	CDKL1
CDKL2	59	CDKL1
CDKL3	56	CDKL1
CDKL4	80	CDKL1
CDKL5	90	CDKL5
CHED	94	CDK13
CHK1	70	CHEK1
CHK2	62	CHEK2
CK1A	87	CSNK1A1
CK1A2	90	CSNK1A1
CK1D	98	CSNK1DA
CK1D	98	CSNK1DB
CK1E	97	CSNK1E
CK1G1	93	CSNK1G1
CK1G2	96	CSNK1G2A
	93	CSNK1G2B
CK1G3	92	CSNK1G1
CK2a1	98	CK2A1
CK2a1	98	ZGC86598

Human kinase	% identity	Zebrafish kinase
CK2a1	98	CSNK2A1
	91	CSNK2A2B
	89	CK2A2A
CK2a2	89	CSNK2A2A
	83	STK35
CLIK1	81	STK35L
	48	SICH211 63O20.7
CLIK1L	84	PDIK1L
CLK1	69	CLK4B
CLK2	88	CLK2A
CLK2	85	CLK2B
CLK3	68	CLK2A
CLK4	70	CLK4B
CLK4	68	CLK4A
СОТ	39	CHUK
CRIK	55	LATS1
CRK7	95	CDK12
CSK	89	CSK
СТК	60	CSK
CYGD	60	GC3
	77	GC2
CYGF	72	GUCY2F
	61	GC3
DAPK1	88	DAPK1
	82	DAPK2
DARK2	82	DAPK2A
	79	DAPK2B
	94	DAPK3
DAPK3	55	SIDKEY- 240H12.4
DCLK1	91	DCKL1B
DCLKI	91	DCLK1A
DCLK2	83	DCLK2A
DCLKZ	83	DCLK2B

Human kinase	% identity	Zebrafish kinase
DCLK2	83	DCLK2C
DCLK2 DCLK3	54	DCLK2C DCLK3
DDR1	46	NTRK2B
DDR2	47	ABL2
DLK	92	MAP3K12
DMPK1	78	SICH211- 89P3.3
DMPK2	79	CDC42BPAA
	83	STK17A
DRAK1	75	ZGC153952
	67	STK17B
DRAK2	71	STK17A
Dusty	89	DSTYK
	99	DYRK1AA
DYRK1A	99	DYRK1AB
	97	DYRK1B
DYRK1B	96	DYRK1B
DUDUA	97	DYRK2
DYRK2	89	ZGC172180
DYRK3	43	SIDKEY-
_		238N5.3
DYRK4	75	DYRK4
EGFR	84	ERBB2
EPHA1	66	EPHB4A
EPHA10	57	EPHA4B
EPHA2	78	EPHA2A
EPHA3	89	EPHA4B
	86	EK1
EPHA4	96	EPHA4A
EPHA5	89	EPHA7
EPHA6	88	EPHA4A
EPHA7	97	EPHA7
EPHA8	80	EPHA7
EPHB1	91	EPHB2B
EPHB2	97	EPHB2B

Table S1 cont	inued
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Human kinase	% identity	Zebrafish kinase
EPHB3	87	EPHB3A
	79	EPHA2A
	88	EPHB4A
EPHB4	88	EPHB4B
EPHB6	55	EPHB3A
ERBB2	90	ERBB2
ERBB3	75	ERBB3A
EKDDJ	70	ERBB3B
ERBB4	95	ERBB4A
Erk1	89	MAPK3
Erk2	96	MAPK1
EIK2	92	MAPK3
Erk3	90	MAPK6
Erk4	68	MAPK4
Erk5	90	MAPK7
Erk7	77	MAPK15
FAK	93	PTK2.1
FAM20A	24	MINK1
FAM20B	18	OXSR1B
FAM20C	26	PIM1
FER	70	FES
FES	76	FES
FGFR1	91	FGFR1B
TOTKI	88	FGFR1A
FGFR2	94	FGFR2
FGFR3	82	FGFR4
FGFR4	77	FGFR2
FGR	83	YRK
FJB1	27	PINK1
FLT1	70	FLT1
FLT3	54	KITB
FLT4	72	FLT4
FMS	68	CSF1RA
FRK	54	PTK6B

Human kinase	% identity	Zebrafish kinase
Fused	84	STK36
FYN	95	FYNB
GAK	84	GAK
GCK	75	MAP4K6
GCN2	67	EIF2AK4
GCN2A	31	MARK1
GPRK4	94	GRK4
GPRK5	74	GRK5
GPRK6	91	GRK5L
CDDVZ	80	GRK7A
GPRK7	76	GRK7B
GGYIG	91	GSK3AA
GSK3A	91	GSK3AB
	98	GSK3B
GSK3B	93	SIDKEYP-
		80C12.7 SIDKEYP-
Haspin	65	26A9.2
HCK	82	LYN
HGK	91	TNIKA
HH498	94	TNNI3K
HIPK1	92	HIPK1A
HIPK2	98	HIPK2
LUDV2	92	HIPK3B
HIPK3	94	HIPK3A
HIPK4	55	HIPK3A
HPK1	68	MAP4K5
HRI	45	EIF2AK1
HSER	47	ERBB2
	77	HUNK
	40	SICH73- 381F5.2
HUNK	40	SIDKEY-93L1.4
	40	SIDKEY-93L1.6
	38	SIDKEY- 183G16.2
ICK	87	MAK

Human kinase	% identity	Zebrafish kinase
IGF1R	89	IGF1RA
	89	IGF1RB
IKKa	75	CHUK
IKKb	79	IKBKB
IKKe	77	IKBKE
ILK	86	ILK
INSR	91	INSRA
IINSK	91	INSRB
IRAK1	63	IRAK1
IRAK2	48	IRAK1
IRAK3	52	IRAK1
IRAK4	54	IRAK4
IRE1	88	ERN1
IKEI	74	ERN2
IRE2	65	ERN1
IRR	79	INSRB
ITK	66	ITK
JAK1-P	70	JAK1-P
JAK1	77	JAK1
JAK2-P	83	JAK2B-P
JAK2	88	JAK2B
JAK2	84	JAK2A
JAK3	68	JAK2B
JAK3-P	60	JAK3-P
JNK1	95	MAPK8A
JNK2	92	MAPK9
INUZ2	99	MAPK10
JNK3	96	MAPK8B
KDD	74	KDRL
KDR	67	KDR
	93	MAP4K5
KHS1	77	MAP4K2L
	77	MAP4K2
KHS2	93	MAP4K3A

Human kinase	% identity	Zebrafish kinase
KUSO	92	MAP4K3B
KHS2	78	MAP4K6
KIS	71	UHMK1
ИТ	71	KITB
KIT	69	KITA
KSR1	77	KSR2
KSR2	88	KSR2
LATS1	96	LATS1
LATS2	93	LATS2
LCK	78	LCK
LIMK1	94	LIMK1A
LIMK2	87	LIMK2
LKB1	96	STK11
LMR1	46	PIM3
LMR2	51	NEK4
LMR3	40	ACVRL1
LOK	76	SLKA
LRRK1	44	LRRK2
LRRK2	72	LRRK2
LTK	45	ACVR2AB
LYN	89	LYN
LZK	86	MAP3K12
MAK	92	MAK
MAP2K1	94	MAP2K1
MAP2K2	96	MAP2K2A
WIAF2K2	93	MAP2K2B
MAP2K3	60	РОМК
MAP2K4	96	MAP2K4A
MAP2K5	89	MAP2K5
MAP2K6	91	MAP2K6
MAP2K7	88	MAP2K7
MAP3K1	93	MAP3K1
MAP3K2	94	MAP3K2
MAP3K3	95	MAP3K3

Table S1 continued..

Human kinase	% identity	Zebrafish kinase
MAP3K4	99	MAP3K4
	95	MAP3K5
MAP3K5	91	SICH211- 1I11.3
	89	MAP3K15
MAP3K6	73	SICH211- 1I11.3
MAP3K7	81	MAP3K15
	88	MAPKAPK2A
MAPKAPK2	73	МАРКАРК2В
МАРКАРКЗ	78	МАРКАРК3
MAPKAPK5	92	MAPKAPK5
MARK1	39	SIDKEY- 206F10.6
MAKKI	38	SIDKEY- 83M22.16
MARK2	98	MARK2A
MARK2	96	MARK2B
	97	MARK3B
MARK3	96	MARK1
MARKS	93	MARK3
	93	MARK3A
MARK4	95	SIDKEY- 31M14.7
	93	MARK4A
MAST1	97	MAST1A
MASTI	96	MAST1B
MAST2	95	MAST2
MAST3	94	MAST1A
MAST4	96	MAST4
MASTL	86	MASTL
MEKK15	75	MAP3K19
MELK	80	MELK
MER	62	TYRO3
MET	89	MET
MINK	92	TNIKB
MISR2	48	CHEK1

Human kinase	% identity	Zebrafish kinase
MLK1	26	PAK7
MLKI	67	PIM3
MLK2	67	PIM3
MLK3	67	PIM3
MLK4	67	PIM3
MLKL	37	TAOK1A
MNK1	83	MKNK1
NOWA	84	MKNK2A
MNK2	83	MKNK2B
MOK	43	CK2A1
MOS	60	MOS
MSK1-P	81	RPS6KA5-P
MPSK1	65	STK16
MSK2	82	RPS6KA4
MSSK1	81	SRPK1A
MST1	95	STK3
MRCKA	95	SICH211- 89P3.3
_	93	CDC42BPAA
MRCKB	90	CDC42BPB
MSK1	89	RPS6KA4
MSKI	89	RPS6KA5
	72	RPS6KA4-P
MSK2-P	39	SIDKEYP- 67E1.4
	37	SIDKEY- 155D18.1
MST2	99	STK3
	97	STK24B
MST3	93	STK24A
	90	STK25A
MST4	93	MST4
	93	STK26
MUSK	80	MUSK
MYO3A	83	MYO3A
MYO3B	80	MYO3B

Human kinase	% identity	Zebrafish kinase
MYT1	50	PKMYT1
NDR1	98	STK38A
NDKI	96	STK38B
NDR2	95	STK38L
	89	NEK1
	56	SIDKEY- 192D15.1
NEK1	52	SIDKEY- 201L21.1
	51	SIDKEY- 125E8.1
	48	SIDKEY- 192D15.2
NEK10	80	NEK10
NEK11	57	PIM3
NEK2	82	NEK2
NEWS	42	ZGC113355
NEK3	42	NEK12
NEK4	86	NEK4
NEK5	74	NEK1
NEK6	87	NEK6
NEK7	89	NEK7
NEK8	85	NEK8
NEK9	47	NEK1
NUZ	56	SICH211- 253B1.4
NIK	54	SIDKEY- 260C8.4
	65	SICH211- 22D5.2
NIM1	40	SIDKEY- 183G16.1
	100	NLK2
NLK	92	NLK1
NME1A	21	SIK2A
NME1B	21	JAK1
NME3	20	PRKD3
NME4	21	JAK1
NME5	21	GUCY2F

NME6 21 POMK NME7a 18 STK10 NRBP1 95 NRBP1 NRBP2 74 NRBP1 NRBP2 74 NRBP1 NRBP2 74 NRBP1 NRK 60 TNIKB 92 NUAK1A 91 NuAK1 69 SICH211- 117C9.1 39 SIDKEY- 197D18.2 39 39 SIDKEY- 197D18.2 39 39 SIDKEY- 197D18.1 24 24 PXK NUAK2 OBSCN-P 57 OBSCN-P OBSCN 50 SICH211- 195M20.1 0SR1 94 OXSR1A 0SR1 92 OXSR1A 0SR1 92 OXSR1A 0SR1 92 OXSR1A 92 OXSR1A 92 0SR1 92 OXSR1A 938 95 MAPK14B 938 95 MAPK13 938 MAPK13<	Human kinase	% identity	Zebrafish kinase
NRBP1 95 NRBP1 NRBP2 74 NRBP1 NRK 60 TNIKB 92 NUAK1A 91 NUAKIB 69 SICH211- 117C9.1 1 197D18.2 39 39 SIDKEY- 197D18.2 39 39 SIDKEY- 197D18.1 24 24 PXK NUAK2 OBSCN-P 57 OBSCN-P OBSCN 50 SICH211- 195M20.1 OBSCN 50 SICH211- 195M20.1 OSR1 94 OXSR1A 92 OXSR1A 92 OSR1 92 OXSR1A 92 OXSR1A 92 OSR1 94 OXSR1A 92 OXSR1A 92 938a 96 MAPK14A 938b 91 MAPK11 p38d 78 MAPK13 69 ZGC17175 98 PA0056K 84 RPS6KB1B PAK1 98	NME6	21	РОМК
NRBP2 74 NRBP1 NRK 60 TNIKB 92 NUAK1A 91 NUAK1B 69 SICH211- 117C9.1 139 SIDKEY- 197D18.2 39 SIDKEY- 197D18.2 39 SIDKEY- 197D18.1 24 PXK NuAK2 80 OBSCN-P 57 OBSCN 50 SICH211- 195M20.1 0SR1 94 0SR1 92 0SR1 94 0SR1 92 0SR1 94 0SR1 92 0SR1 92 0SR1 92 0SR1 92 0SR1 93 95 MAPK14B p38a 95 938 91 938 91 938 93 938 94 938 95 938 93 94 RPS6KB1B <	NME7a	18	STK10
NRK 60 TNIKB NRK 60 TNIKB 92 NUAK1A 91 NUAK1B 69 SICH211- 117C9.1 139 SIDKEY- 197D18.2 39 SIDKEY- 197D18.2 39 SIDKEY- 197D18.1 24 PXK NuAK2 80 NUAK2 OBSCN-P 57 OBSCN-P 0BSCN 50 SICH211- 195M20.1 0BSCN 94 OXSR1A 0SR1 94 OXSR1A 92 OXSR1A 92 0SR1 92 OXSR1A 938 91 MAPK14B p38a 95 MAPK14A p38d 78 MAPK13 69 ZGC171775 98 p38g 79 MAPK13 69 ZGC171775 98 PA056K 84 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A <td< td=""><td>NRBP1</td><td>95</td><td>NRBP1</td></td<>	NRBP1	95	NRBP1
92 NUAK1A 91 NUAK1B 69 SICH211- 117C9.1 139 SIDKEY- 197D18.2 39 SIDKEY- 197D18.2 39 SIDKEY- 197D18.1 24 PXK NuAK2 80 0BSCN-P 57 0BSCN 50 0BSCN 50 0BSCN 94 0SR1 92 94 OXSR1A 92 OXSR1A 93 SICH211- 195M20.1 0BSCN 50 SICH211- 195M20.1 0SR1 94 OXSR1A 92 OXSR1A 92 0SR1 92 OXSR1A 92 OXSR1A 93 938a 91 MAPK14A 938b 91 MAPK11 p38d 78 MAPK12A 83 MAPK12A 81 938g 79 MAPK13 69 ZGC171775 98 PAK1 98 </td <td>NRBP2</td> <td>74</td> <td>NRBP1</td>	NRBP2	74	NRBP1
NuAK1 91 NUAK1B 91 NUAK1B 69 SICH211- 117C9.1 39 SIDKEY- 197D18.2 39 SIDKEY- 197D18.2 39 SIDKEY- 197D18.1 24 PXK NuAK2 80 NUAK2 OBSCN-P 57 OBSCN-P OBSCN 50 SICH211- 195M20.1 OBSCN 94 OXSR1A OSR1 92 OXSR1A P38a 96 MAPK14B p38a 91 MAPK14A p38b 91 MAPK13 p38d 78 MAPK13 p38d 78 MAPK12A p38g 69 ZGC171775 p38g 98 RPS6KB1B P70S6K 94 RPS6KB1A P70S6K 94 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4 <td>NRK</td> <td>60</td> <td>TNIKB</td>	NRK	60	TNIKB
NuAK1 69 SICH211- 117C9.1 39 SIDKEY- 197D18.2 39 SIDKEY- 197D18.1 24 PXK NuAK2 80 NUAK2 OBSCN-P 57 OBSCN-P OBSCN 50 SICH211- 195M20.1 OBSCN 94 OXSR1A OSR1 92 OXSR1A p38a 95 MAPK14B p38a 95 MAPK14 p38b 91 MAPK13 p38d 78 MAPK13 p38g 79 MAPK13 p38g 79 MAPK13 p70S6K 94 RPS6KB1B P70S6K 94 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4		92	NUAK1A
NuAK1 69 117C9.1 39 SIDKEY- 197D18.2 39 SIDKEY- 197D18.1 24 PXK NuAK2 80 NUAK2 OBSCN-P 57 OBSCN-P OBSCN 50 SICH211- 195M20.1 OBSCN 94 OXSR1A OSR1 92 OXSR1A 938 96 MAPK14B p38a 96 MAPK14A p38a 91 MAPK11 p38d 78 MAPK13 p38g 78 MAPK13 p38g 79 MAPK13 p38g 98 RPS6KB1B p70S6K 98 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4		91	NUAK1B
39 SIDKE1- 197D18.2 39 SIDKEY- 197D18.1 24 PXK 24 PXK 0BSCN-P 57 OBSCN-P 0BSCN 50 SICH211- 195M20.1 0BSCN 50 SICH211- 195M20.1 0SR1 94 OXSR1A 0SR1 92 OXSR1B p38a 96 MAPK14B p38a 95 MAPK14 p38b 91 MAPK13 p38d 78 MAPK13 p38g 79 MAPK13 p38g 79 MAPK13 69 ZGC171775 98 P70S6K 98 RPS6KB1B PAK1 98 PAK1 PAK1 98 PAK1 PAK1 96 PAK2A PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4		69	
39 197D18.1 24 PXK NuAK2 80 NUAK2 OBSCN-P 57 OBSCN-P OBSCN 50 SICH211- 195M20.1 OBSCN 94 OXSR1A OSR1 92 OXSR1A P38a 96 MAPK14B p38a 95 MAPK14A p38b 91 MAPK11 p38d 78 MAPK13 p38g 83 MAPK12B p38g 79 MAPK13 69 ZGC171775 98 P70S6K 94 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4	NuAK1	39	
NuAK2 80 NUAK2 OBSCN-P 57 OBSCN-P OBSCN 50 SICH211- 195M20.1 OBSCN 94 OXSR1A OSR1 92 OXSR1B P38a 96 MAPK14B P38a 95 MAPK14A P38b 91 MAPK11 P38d 78 MAPK13 P38g 78 MAPK12A P38g 91 MAPK13 P38g 91 MAPK13 P38g 98 MAPK13 P38g 79 MAPK13 69 ZGC17175 SGC17175 P70S6K 98 RPS6KB1B P4K1 98 PAK1 PAK2 96 PAK2A PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4		39	
OBSCN-P 57 OBSCN-P OBSCN 50 SICH211- 195M20.1 OSR1 94 OXSR1A 92 OXSR1B p38a 96 MAPK14B p38a 95 MAPK14A p38b 91 MAPK11 p38d 78 MAPK13 p38d 78 MAPK13 p38g 78 MAPK13 p38g 78 MAPK13 p38g 79 MAPK13 p38g 79 MAPK13 p70 MAPK13 69 p70S6K 98 RPS6KB1B P70S6K 94 RPS6KB1A P70S6KB 84 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4		24	РХК
OBSCN 50 SICH211- 195M20.1 OSR1 94 OXSR1A 92 OXSR1B p38a 96 MAPK14B p38a 95 MAPK14A p38b 91 MAPK11 p38d 78 MAPK13 p38d 78 MAPK13 p38g 83 MAPK12A p38g 79 MAPK13 p38g 79 MAPK13 p38g 98 RPS6KB1B P70S6K 98 RPS6KB1B PAK1 98 PAS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4	NuAK2	80	NUAK2
OBSCN 50 195M20.1 OSR1 94 OXSR1A 92 OXSR1B p38a 96 MAPK14B p38a 95 MAPK14A p38b 91 MAPK11 p38d 78 MAPK13 p38d 78 MAPK12A p38g 83 MAPK12A p38g 79 MAPK13 p38g 79 MAPK13 p38g 79 MAPK13 p70S6K 98 RPS6KB1B PAK1 98 PAS6KB1A PAK2 96 PAK2A 96 PAK2A 96 PAK3 97 PAK1 PAK4 93 PAK4	OBSCN-P	57	OBSCN-P
OSR1 92 OXSR1B p38a 96 MAPK14B p38a 95 MAPK14A p38b 91 MAPK11 p38d 78 MAPK13 p38d 78 MAPK13 p38g 83 MAPK12A p38g 69 ZGC17175 P70S6K 98 RPS6KB1B P70S6KB 84 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4	OBSCN	50	
92 OXSR1B p38a 96 MAPK14B p38a 95 MAPK14A p38b 91 MAPK11 p38d 78 MAPK13 p38d 78 MAPK12A p38d 78 MAPK13 p38g 83 MAPK12B p38g 79 MAPK13 69 ZGC171775 P70S6K 98 RPS6KB1B PAK1 98 PAS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A 96 PAK2A 96 PAK3 97 PAK1 PAK4 93 PAK4	0001	94	OXSR1A
p38a 95 MAPK14A p38b 91 MAPK11 p38d 78 MAPK13 p38d 78 MAPK13 p38d 78 MAPK13 p38d 78 MAPK13 p38g 83 MAPK12A p38g 69 ZGC17175 P70S6K 98 RPS6KB1B P70S6KB 84 RPS6KB1A PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4	OSRI	92	OXSR1B
95 MAPK14A p38b 91 MAPK11 p38d 78 MAPK13 p38d 78 MAPK13 p38d 78 MAPK13 p38d 78 MAPK12A p38g 83 MAPK12A p38g 69 ZGC17175 P70S6K 98 RPS6KB1B P70S6KB 84 RPS6KB1A P70S6KB 84 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4	20	96	MAPK14B
p38d 78 MAPK13 p38g 78 MAPK12A 83 MAPK12B p38g 81 MAPK12B p38g 79 MAPK13 69 ZGC171775 P70S6K 98 RPS6KB1B P70S6KB 84 RPS6KB1A P70S6KB 84 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4	p38a	95	MAPK14A
83 MAPK12A p38g 81 MAPK12B 79 MAPK13 69 ZGC171775 P70S6K 98 RPS6KB1B P70S6KB 84 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4	p38b	91	MAPK11
81 MAPK12B 79 MAPK13 69 ZGC171775 P70S6K 98 RPS6KB1B P70S6K 94 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4	p38d	78	MAPK13
p38g 79 MAPK13 69 ZGC171775 P70S6K 98 RPS6KB1B P70S6K 94 RPS6KB1A P70S6KB 84 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4		83	MAPK12A
79 MAPK13 69 ZGC171775 P70S6K 98 RPS6KB1B 94 RPS6KB1A P70S6KB 84 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4	20	81	MAPK12B
98 RPS6KB1B 94 RPS6KB1A 94 RPS6KB1A P70S6KB 84 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4	p38g	79	MAPK13
P70S6K 94 RPS6KB1A P70S6KB 84 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4		69	ZGC171775
94 RPS6KB1A P70S6KB 84 RPS6KB1B PAK1 98 PAK1 PAK2 96 PAK2A PAK3 97 PAK1 PAK4 93 PAK4		98	RPS6KB1B
PAK1 98 PAK1 PAK2 96 PAK2A 96 PAK2B 96 PAK3 97 PAK1 PAK4 93 PAK4	P70S6K	94	RPS6KB1A
PAK2 96 PAK2A 96 PAK2B 96 PAK2B PAK3 97 PAK1 PAK4 93 PAK4	P70S6KB	84	RPS6KB1B
PAK2 96 PAK2B PAK3 97 PAK1 PAK4 93 PAK4	PAK1	98	PAK1
96 PAK2B PAK3 97 PAK1 PAK4 93 PAK4		96	
PAK4 93 PAK4	PAK2	96	PAK2B
PAK4 93 PAK4	PAK3	97	PAK1
PAK4		93	PAK4
	PAK4		

Table S1	continued
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Human kinase	% identity	Zebrafish kinase
PAK5	91	PAK4
PAK6	89	PAK6B
PAN3	26	DCKL1B
	44	SIDKEY-
	44	37M8.3
	40	SIDKEY-
		37M8.13
	40	SIDKEY-
		11F7.5 SIDKEY-
	40	278K10.4
PASK		SIDKEY-
	39	37M8.14
	20	SIDKEY-
	38	211E20.1
	38	SICH211-
	50	57M13.8
	38	SICH211-
		249H16.8
PBK	70	PBK
PCTAIRE1	93	CDK16
PCTAIRE2	96	CDK16
PCTAIRE3	90	CDK16
PDGFRA	58	PDGFRB
PDGFRB	68	PDGFRB
PDK1	88	PDPK1B
IDKI	83	PDPK1A
PEK	57	EIF2AK3
PFTAIRE1	99	CDK14
PFTAIRE2	85	CDK15
PHKG1	78	PHKG1A
FIIKOI	76	PHKG1B
PHKG2	75	PHKG2
PIK3R4	44	CHEK1
	76	PIM1
	54	SICH211- 231M1.1
PIM1	53	SICH211- 193E5.3
	53	SIDKEY- 45J10.1

Table	S 1	continued

Human kinase	% identity	Zebrafish kinase
	53	SIDKEY-
		45D16.8
		SICH211-
	52	194C3.4
		SIDKEY-
	52	101N15.3
	52	SIDKEY-11E6.2
	52	SIDKEY-11E6.4
	52	SIDKEY-
	52	278G4.6
	52	SICH211-
	52	244K5.5
	52	SIDKEY-
	32	10F21.3
	51	SIDKEY-
	51	278G4.4
	47	SICH211-
	47	155D24.5
	46	SICH211-
	40	113J14.2
	46	SICH211-
	40	222N4.5
DD (1	46	SICH73-
PIM1	40	272H16.1
	45	SICH73-
	45	272H16.3
	44	SIDKEYP-
	44	104H9.7
	44	SICH211-
		126G16.5
	44	SIDKEY-
	-+-+	258F14.4
	44	SICH211-
		126G16.2
	44	SIDKEY-
		205D4.2
	43	SIDKEY-
	-15	58B18.10
	41	SICH211-
	71	255L14.1
	41	SICH211-
	71	255L14.3
	41	SICH211-
	41	255L14.5
	/1	SIDKEY-
	41	236A14.13
	40	SICH211-
	40	165A10.4

Zebrafish			
Human kinase	% identity	kinase	
	40	SIDKEY-	
	40	40H17.6	
	10	SICH211-	
	40	165A10.5	
	20	SIDKEY-	
	38	236A14.6	
	38	SICH211-	
		196G2.8	
DIM1		SICH211-	
PIM1	38	119D14.4	
		SICH211-	
	38	198E20.11	
		SIDKEYP-	
	37	100A1.5	
		SICH211-	
	34	196G2.7	
	22	SICH211-	
	33	168M18.3	
	45	SIDKEY-	
	43	58B18.9	
	44	SIDKEY-	
		31M5.5	
	44	SIDKEY-	
		31M5.7	
	43	SIDKEY-	
		31M5.3	
	41 41 40 39 39	SIDKEY-	
		149I17.9	
		SICH211-	
PIM2		10J20.5 SICH211-	
		10J20.1	
		SIDKEYP-	
		110E4.3	
		SICH211-	
		215M21.11	
	20	SICH211-	
	39	176G13.9	
	20	SICH211-	
	39	237D5.4	
	26	SICH211-	
	36	57M13.10	
	74	PIM3	
	59	PIM2	
PIM3	54	SIDKEY-	
		278G4.9	
	54	SIDKEY-	
	J4	193D10.6	

Table S1 continued..

H	0/ 10-0414-0	Zebrafish
Human kinase	% identity	kinase
	5 4	SIDKEY-
	54	278G4.1
		SICH211-
	53	193E5.4
		SIDKEY-
	53	43A14.3
		SIDKEY-
	53	43A14.4
		SICH211-
	53	207C6.10
	53	SIDKEY-
		193D10.8
	53	SICH211-
		231M1
	53	SIDKEY-
	55	278G4.3
	53	SICH211-
	55	244K5.3
	52	SIDKEY-
	53	278G4.8
	50	SICH211-
	53	231M1.3
		SIDKEY-
	53	278G4.2
		SIDKEY-
PIM3	53	11E6.5
		SIDKEY-
	53	
		11E6.3
	53	SICH211-
		231M1.4
	53	SIDKEY-
		193D10.2
	53	SIDKEY-
	55	193D10.7
	53	SIDKEY-
	55	193D10.5
	52	SIDKEY-
	53	278G4.7
	52	SIDKEY-
	53	278G4.10
		SICH211-
	53	244K5.4
		SICH211-
	53	231M1.5
		SICH211-
	52	
		207C6.9
	52	SIDKEY-
		45D16.9
	52	SICH211-
	52	244K5.2

	Zebrafish				
Human kinase	% identity	kinase			
		SIDKEY-			
	52	193D10.11			
		SIDKEY-			
	52	193D10.3			
		SIDKEY-			
	52	265A11.5			
	51	SIDKEY-			
	51	278G4.5			
	51	SIDKEY-			
		11E6.1			
	51	SIDKEY-			
		236A14.11 SIDKEY-			
	51	236A14.10			
		SIDKEY-			
	51	236A14.5			
		SIDKEY-			
	51	236A14.5			
	51	SIDKEY-			
	51	236A14.11			
	51	SIDKEY-1P9.2			
	50	SIDKEY-			
	50	43A14.5			
	50	SIDKEY-			
PIM3	30	236A14.4			
	50	SIDKEY-			
		236A14.4			
	46	SIDKEY-			
		58B18.3			
	45	SICH73-			
		129A22.11			
	45	SICH211-			
		214C11.9 SICH211-			
	45	196N4.2			
		SICH211-			
	45	196N4.1			
	15	SIDKEY-			
	45	179A6.3			
	44	SIDKEY-			
	44	179A6.1			
	44	SICH211-			
		196N4.3			
	44	SICH211-			
	44	214C11.6			
		SIDKEY- 179A6.2			
		SIDKEY-			
	44	43B14.9			
	L				

Human kinase	% identity	Zebrafish kinase
	44	SICH211-
		270P4.3
	44	SIDKEY-
		149I17.8
	44	SICH211-
	44	254P10.3
	44	SIDKEY-
	44	208K22.2
	12	SIDKEY-
	43	43B14.7
	43	SIDKEY-
	43	43B14.6
		SICH211-
	43	126G16.3
		SICH211-
	43	126G16.4
		SIDKEY-
	43	205012.2
		SIDKEY-
	43	58B18.6
		SICH211-
	43	215M21.17
	43	SIDKEY- 14A7.4
		SICH211-
PIM3	43	214C11.3
	43	SIDKEY-
		108D22.5
		SIDKEY-
		33C12.6
		SIDKEY-
	42	8403.91
	42	SIDKEY-
		84O3.11
		SIDKEY-
	42	84O3.13
		SIDKEY-
	42	84O3.10
	42	SIDKEY-
		84O3.15
-	42	SIDKEY-
		33C12.8
	42	SIDKEY-
		112G5.2
	42	SIDKEY-
	42	10B15.18
		SIDKEY-
		112G5.7
	42	SIDKEY-
		33C12.7

Human kinase	% identity	Zebrafish
numan kinase	76 Identity	kinase
	42	SIDKEY-
		8403.12
	42	SIDKEY-
		84O3.14
	42	SIDKEY-
		112G5.5
	42	SIDKEY-
	12	19P15.5
	39	SICH211-
		57M13.6
	39	SICH211-
		57M13.3
	39	SICH211-
		57M13.5
	39	SICH211-
		57M13.7 SICH211-
	39	147H1.4
PIM3		SIDKEYP-
	39	67E1.6
		SICH211-
	35	196N4.7
		SICH211-
	35	214C11.2
	34	SIDKEY-
		222B8.5
	32	SICH211-
		214C11.4
	32	SICH211-
		196N4.8
	32	SICH211-
		214C11.8
	32	SICH211-
		196N4.5
	31	SICH211-
		214C11.7
PINK1	67	PINK1
DKACA	95	PRKACAA
PKACA	91	PRKACAB
	94	PRKACBB
PKACB	90	PRKACBA
PKACG	80	PRKACAB
РКСА	91	PRKCA
PKCB	93	PRKCBB
FNUD	91	PRKCBA

% identity	Zebrafish kinase
87	PRKCDA
87	PRKCDB
97	PRKCEA
96	PRKCEB
77	PRKCG
80	PRKCHB
80	PRKCHA
99	PRKCI
88	PRKCQ
96	PRKCZ
96	PRKD1
39	SIDKEY- 211E20.8
30	SIDKEY-
	211E20.9
73	ZGC175248
94	PRKD1
99	PRKG1B
96	PRKG1A
61	SIDKEY- 121J17.5
80	PRKG2
86	PKN1A
94	PKN2
94	ZGC153916
86	PKN1A
75	PKN3
70	PKN1A
53	PKZ
47	EIF2AK2
81	PLK1
89	PLK2A
81	PLK2B
82	PLK3
83	PLK4
82	PRKX
	87 87 97 96 77 80 80 99 88 96 39 39 39 39 39 73 94 99 96 61 80 86 94 94 94 95 70 53 47 81 89 81 82 83

Human kinase	% identity	Zebrafish
PRKY	82	kinase PRKX
TKKT	99	PRPF4BB
PRP4	95	PRPF4BA
PRPK	64	TP53RK
FKFK	-	
PSKH1	91	PSKH1 SICH211-
	66	27E6.1
PSKH2	74	PSKH1
PYK2	72	PTK2BB
OW	92	SIK2B
QIK	86	SIK2A
QSK	93	ZGC66101
	95	RAF1A
RAF1	94	RAF1B
RET	88	RET
DUOV	86	GRK1A
RHOK	82	GRK1B
RIPK1	68	RIPK1L
RIPK2	68	RIPK2
RIPK3	57	PIM3
RNAseL	52	CHEK1
ROCK1	98	ROCK1
	93	ROCK2A
ROCK2	82	ROCK2B
RON	65	MET
ROR1	43	RYK
ROR2	43	NTRK2B
ROS	48	ABL2
RSK1	96	RPS6KA1
RSK1-P	87	RPS6KA1-P
RSK2-P	89	RPS6KA3A-P
DGW2	96	RPS6KA3A
RSK2	95	RPS6KA3B
RSK3	92	RPS6KA3A
RSK3-P	87	RPS6KA2-P

Human kinase	% identity	Zebrafish kinase
RSK4-P	86	RPS6KA6-P
RSK4	96	RPS6KA6
RSKL1	48	CHEK1
RSKL2	48	CHEK1
RYK	89	RYK
	78	SICH211- 183D21.3
	69	BSK146A
SBK	68	BSK146B
	48	SICH211- 171H4.3
	47	SBK3
SCYL1	48	CAMKVL
SCYL2	89	SICH211- 244B2.1
SCYL3	73	SCYL3
SgK069	61	SIDKEY- 8E10.3
SgK071	50	CHEK1
	76	MYLK4A
SGK085	74	MYLK4B
	67	MYLK2
	95	SGK1
SGK1	88	SICH211- 195B13.1
	72	SGK2B
SgK110	50	SICH211- 183D21.3
SgK196	60	POMK
SGK2	86	SGK2A
SgK223	20	PRKD3
SgK269	25	PRKD3
SGK288	64	ANKK1
SGK3	88	SGK3
SgK307	50	РОМК
SgK396	33	MAP3K12
SgK493	34	AURKA
SGK494	59	SGK494A

Human kinase	% identity	Zebrafish kinase
SGK494	53	SGK494B
SGK495	30	PTK6B
	91	SIK1
SIK	36	SICH211- 57M13.9
SKMLCK	66	MYLK4B
	88	SLKA
SLK	87	SLKB
	70	SIDKEY- 81J8.6
Slob	31	CDK13
	81	MYLKA
SMMLCK	79	MYLKB
	62	SIDKEY- 194J22.6
SNRK	98	SNRKA
SINKK	85	SNRKB
SPEG	57	SICH211- 195M20.1
SPEG-P	67	SICH211- 195M20.1
	62	SPEG
SRC	95	SRC
SRM	53	FYNB
SRPK1	91	SRPK1A
SKEKI	88	SRPK1B
SRPK2	85	SRPK1A
SSTK	55	TSSK6
STK33	70	STK33
STLK3	87	OXSR1A
STLK5	72	STRADA
STLK6	47	STRADA
SURTK106	41	PIM3
SYK	80	SYK
TAK1	90	MAP3K7
TAO1	96	TAOK1A
TAO2	92	TAOK2A

Human kinase	% identity	Zebrafish kinase
TAO2	92	TAOK2B
TAO3	88	TAOK3A
TBCK	62	TBCK
TBK1	88	TBK1
TEC	75	TEC
TESK1	52	ZGC113162
TESK2	85	TESK2
TGFBR1	99	TGFBR1A
TOPBRI	99	TGFBR1B
	88	TGFBR2
TGFBR2	52	SIDKEY- 101K6.5
TIE1	77	TEK
TIE2	90	TEK
TLK1	86	TLK1B
ILKI	86	TLK1A
TLK2	87	TLK2
	97	TNIKB
TNIK	94	TNIKA
	61	MINK1
TNK1	49	TNK1
TRAD	57	TRIOA
TRB1	55	TRIB1
TRB2	58	TRIB3
TRB3	49	TRIB3
TRIO	70	TRIOA
IKIO	67	TRIOB
TRKA	73	NTRK2B
TRKB	90	NTRK2B
TRKC	95	NTRK3A
	40	SICH211- 215M21.25
TSSK1	40	SICH211- 215M21.23
	40	SICH211- 220I15.5

Hamon binoso	0/ :	Zebrafish
Human kinase	% identity	kinase
	40	SICH211-
	40	215M21.21
	40	SICH211-
		220I15.3
	39	SIDKEY-
	39	206F10.7
TSSK1	38	SIDKEY-
155K1	50	217M5.9
	38	SICH211-
	50	10J20.2
	37	SICH211-
	57	215M21.13
	37	SICH211-
		10J20.4
	41	SIDKEY-
TSSK2		14A7.2
	39	SIDKEY-
		197D18.3 SIDKEY-
	39	248F6.3
		SICH211-
	38	160D20.5
		SICH211-
	38	238G23.1
TOOKA	20	SIZFOS-
TSSK3	38	754C12.2
	37	SICH211-
		207C6.4
	37	SICH211-
		202N12.2
	36	SICH211-
		57M13.1
	38	SICH211-
TSSK4		12E13.5
	38	SICH211-
		12E13.6
TTBK1	95	TTBK1B
	86	SIDKEY-
TTBK2		12H9.11
	27	PRKD3
TTK	67	TTK
TTN	74	TTNB
ТХК	68	TEC
TXNDC3	23	PKN1B
TXNDC6	19	CAMKVL
TYK2	58	JAK1

Table S1	continued
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Human kinase	% identity	Zebrafish kinase
TYK2-P	64	TYK2-P
TYRO3	71	TYRO3
ULK1	85	ULK1B
	78	ULK1A
ULK2	90	ULK2
ULK3	79	ULK3
ULK4	39	STK36
VACAMKL	91	CAMKVA
	89	CAMKVB
	70	CAMKVL
VRK1	73	VRK1
VRK2	67	VRK2
VRK3	46	VRK3
Wee1	77	WEE1
	67	WEE2
Wee1B	65	WEE2
Wnk1	94	WNK1A
	94	WNK1B
	86	SIDKEYP- 61G11.1
Wnk2	93	IM7152756
Wnk3	98	IM7152756
Wnk4	83	WNK1A
YANK1	87	STK32A
YANK2	87	STK32A
YANK3	81	STK32A
YES	95	YES1
	89	YRK
YSK1	95	STK25B
ZAK	82	ZAK
ZAP70	71	ZAP70

REFERENCES

 Manning, G., Whyte, D. B., Martinez, R., Hunter, T. and Sudarsanam, S. 2002, Science, 298(5600), 1912-1934.

- Cohen, P. 2002, Nat. Rev. Drug Discov., 1(4), 309-315.
- 3. Hunter, T. 1995, Cell, 80(2), 225-236.
- 4. Hantschel, O. 2015, ACS Chem. Biol., 10(1), 234-245.
- 5. Zhang, C. and Bollag, G. 2010, Curr. Opin. Genet. Dev., 20(1), 79-86.
- Patterson, H., Nibbs, R., McInnes, I. and Siebert, S. 2014, Clin. Exp. Immunol., 176(1), 1-10.
- Fisk, M., Gajendragadkar, P. R., Maki-Petaja, K. M., Wilkinson, I. B. and Cheriyan, J. 2014, Am. J. Cardiovasc. Drugs, 14(3), 155-165.
- Wang, Q., Zorn, J. A. and Kuriyan, J. 2014, Methods Enzymol., 548, 23-67.
- Mulder, N. J., Apweiler, R., Attwood, T. K., 9. Bairoch, A., Bateman, A., Binns, D., Bork, P., Buillard, V., Cerutti, L., Copley, R., Courcelle, E., Das, U., Daugherty, L., Dibley, M., Finn, R., Fleischmann, W., Gough, J., Haft, D., Hulo, N., Hunter, S., Kahn, D., Kanapin, A., Kejariwal, A., Labarga, A., Langendijk-Genevaux, P. S., Lonsdale, D., Lopez, R., Letunic, I., Madera, M., Maslen, J., McAnulla, C., McDowall, J., Mistry, J., Mitchell, A., Nikolskaya, A. N., Orchard, S., Orengo, C., Petryszak, R., Selengut, J. D., Sigrist, C. J., Thomas, P. D., Valentin, F., Wilson, D., Wu, C. H. and Yeats, C. 2007, Nucleic Acids Res., 35, D224-228.
- Hanks, S. K. and Hunter, T. 1995, FASEB J., 9(8), 576-596.
- Stout, T. J., Foster, P. G. and Matthews, D. J. 2004, Curr. Pharm. Des., 10(10), 1069-1082.
- 12. Hanks, S. K. and Quinn, A. M. 1991, Methods Enzymol., 200, 38-62.
- 13. Hunter, T. 2014, Cold Spring Harb. Perspect. Biol., 6(5), a020644.
- Schomburg, I., Chang, A., Hofmann, O., Ebeling, C., Ehrentreich, F. and Schomburg, D. 2002, Trends Biochem. Sci., 27(1), 54-56.
- 15. Manning, G., Plowman, G. D., Hunter, T. and Sudarsanam, S. 2002, Trends Biochem. Sci., 27(10), 514-520.
- 16. Miranda-Saavedra, D. and Barton, G. J. 2007, Proteins, 68(4), 893-914.
- Boudeau, J., Miranda-Saavedra, D., Barton, G. J. and Alessi, D. R. 2006, Trends Cell Biol., 16(9), 443-452.

- Zeqiraj. E. and van Aalten, D. M. 2010, Curr. Opin. Struct. Biol., 20(6), 772-781.
- Paksa, A. and Raz, E. 2015, Curr. Opin. Cell Biol., 36, 80-85.
- 20. Phillips, J. B. and Westerfield, M. 2014, Dis. Model Mech., 7(7), 739-743.
- Gibert, Y., Trengove, M. C. and Ward, A. C. 2013, Curr. Med. Chem., 20(19), 2458-2466.
- 22. Howe, K., Clark, M. D., Torroja, C. F., Torrance, J., Berthelot, C., Muffato, M., Collins, J. E., Humphray, S., McLaren, K., Matthews, L., McLaren, S., Sealy, I., Caccamo, M., Churcher, C., Scott, C., Barrett, J. C., Koch, R., Rauch, G. J., White, S., Chow, W., Kilian, B., Quintais, L. T., Guerra-Assuncao, J. A., Zhou, Y., Gu, Y., Yen, J., Vogel, J. H., Eyre, T., Redmond, S., Banerjee, R., Chi, J., Fu, B., Langley, E., Maguire, S. F., Laird, G. K., Lloyd, D., Kenyon, E., Donaldson, S., Sehra, H., Almeida-King, J., Loveland, J., Trevanion, S., Jones, M., Quail, M., Willey, D., Hunt, A., Burton, J., Sims, S., McLay, K., Plumb, B., Davis, J., Clee, C., Oliver, K., Clark, R., Riddle, C., Elliot, D., Threadgold, G., Harden, G., Ware, D., Begum, S., Mortimore, B., Kerry, G., Heath, P., Phillimore, B., Tracey, A., Corby, N., Dunn, M., Johnson, C., Wood, J., Clark, S., Pelan, S., Griffiths, G., Smith, M., Glithero, R., Howden, P., Barker, N., Lloyd, C., Stevens, C., Harley, J., Holt, K., Panagiotidis, G., Lovell, J., Beasley, H., Henderson, C., Gordon, D., Auger, K., Wright, D., Collins, J., Raisen, C., Dyer, L., Leung, K., Robertson, L., Ambridge, K., Leongamornlert, D., McGuire, S., Gilderthorp, R., Griffiths, C., Manthravadi, D., Nichol, S., Barker, G., Whitehead, S., Kay, M., Brown, J., Murnane, C., Gray, E., Humphries, M., Sycamore, N., Barker, D., Saunders, D., Wallis, J., Babbage, A., Hammond, S., Mashreghi-Mohammadi, M., Barr, L., Martin, S., Wray, P., Ellington, A., Matthews, N., Ellwood, M., Woodmansey, R., Clark, G., Cooper, J., Tromans, A., Grafham, D., Skuce, C., Pandian, R., Andrews, R., Harrison, E., Kimberley, A., Garnett, J., Fosker, N., Hall, R., Garner, P., Kelly, D., Bird, C., Palmer, S., Gehring, I., Berger, A., Dooley, C. M.,

Ersan-Ürün, Z., Eser, C., Geiger, H., Geisler,
M., Karotki, L., Kirn, A., Konantz, J.,
Konantz, M., Oberländer, M., Rudolph-Geiger, S., Teucke, M., Lanz, C., Raddatz,
G., Osoegawa, K., Zhu, B., Rapp, A.,
Widaa, S., Langford, C., Yang, F., Schuster,
S. C., Carter, N. P., Harrow, J., Ning, Z.,
Herrero, J., Searle, S. M., Enright, A.,
Geisler, R., Plasterk, R. H., Lee, C.,
Westerfield, M., de Jong, P. J., Zon, L. I.,
Postlethwait, J. H., Nüsslein-Volhard, C.,
Hubbard, T. J., Roest Crollius, H., Rogers, J.
and Stemple, D. L. 2013, Nature, 496(7446),
498-503.

- 23. Glasauer, S. M. and Neuhauss, S. C. 2014, Mol. Genet. Genomics, 289(6), 1045-1060.
- Kok, F. O., Shin, M., Ni, C. W., Gupta, A., Grosse, A. S., van Impel, A., Kirchmaier, B. C., Peterson-Maduro, J., Kourkoulis, G., Male, I., DeSantis, D. F., Sheppard-Tindell, S., Ebarasi, L., Betsholtz, C., Schulte-Merker, S., Wolfe, S. A. and Lawson, N. D. 2015, Dev. Cell, 32(1), 97-108.
- Cunningham, F., Amode, M. R., Barrell, D., 25. Beal, K., Billis, K., Brent, S., Carvalho-Silva, D., Clapham, P., Coates, G., Fitzgerald, S., Gil, L., Girón, C. G., Gordon, L., Hourlier, T., Hunt, S. E., Janacek, S. H., Johnson, N., Juettemann, T., Kähäri, A. K., Keenan, S., Martin, F. J., Maurel, T., McLaren, W., Murphy, D. N., Nag, R., Overduin, B., Parker, A., Patricio, M., Perry, E., Pignatelli, M., Riat, H. S., Sheppard, D., Taylor, K., Thormann, A., Vullo, A., Wilder, S. P., Zadissa, A., Aken, B. L., Birney, E., Harrow, J., Kinsella, R., Muffato, M., Ruffier, M., Searle, S. M. J., Spudich, G., Trevanion, S. J., Yates, A., Zerbino, D. R. and Flicek, P. 2015, Nucleic Acids Res., 43, D662-D669.
- Howe, D. G., Bradford, Y. M., Conlin, T., Eagle, A. E., Fashena, D., Frazer, K., Knight, J., Mani, P., Martin, R., Moxon, S. A., Paddock, H., Pich, C., Ramachandran, S., Ruef, B. J., Ruzicka, L., Schaper, K., Shao, X., Singer, A., Sprunger, B., Van Slyke, C. E. and Westerfield, M. 2013 Nucleic Acids Res., 41, D854-860.
- Corpet, F. 1988, Nucleic Acids Res., 16(22), 10881-10890.

- McWilliam, H., Li, W., Uludag, M., Squizzato, S., Park, Y. M., Buso, N., Cowley, A. P. and Lopez, R. 2013, Nucleic Acids Res., 41, W597-600.
- Bollag, G., Hirth, P., Tsai, J., Zhang, J., Ibrahim, P. N., Cho, H., Spevak, W., Zhang, C., Zhang, Y., Habets, G., Burton, E. A., Wong, B., Tsang, G., West, B. L., Powell, B., Shellooe, R., Marimuthu, A., Nguyen, H., Zhang, K. Y., Artis, D. R., Schlessinger, J., Su, F., Higgins, B., Iyer, R., D'Andrea, K., Koehler, A., Stumm, M., Lin, P. S., Lee, R. J., Grippo, J., Puzanov, I., Kim, K. B., Ribas, A., McArthur, G. A., Sosman, J. A., Chapman, P. B., Flaherty, K. T., Xu, X., Nathanson, K. L. and Nolop, K. 2010, Nature, 467(7315), 596-599.
- Chrencik, J. E., Patny, A., Leung, I. K., Korniski, B., Emmons, T. L., Hall, T., Weinberg, R. A., Gormley, J. A., Williams, J. M., Day, J. E., Hirsch, J. L., Kiefer, J. R.,

Leone, J. W., Fischer, H. D., Sommers, C. D., Huang, H. C., Jacobsen, E. J., Tenbrink, R. E., Tomasselli, A. G. and Benson, T. E. 2010, J. Mol. Biol., 400(3), 413-433.

- Yang, J., Yan, R., Roy, A., Xu, D., Poisson, J. and Zhang, Y. 2015, Nat. Methods, 12(1), 7-8.
- Tagliabracci, V. S., Pinna, L. A. and Dixon, J. E. 2013, Trends Biochem. Sci., 38(3), 121-130.
- Desvignes, T., Pontarotti, P., Fauvel, C. and Bobe, J. 2009, BMC Evol. Biol., 9, 256.
- 34. Pearce, D. 2003, Cell Physiol. Biochem., 13(1), 13-20.
- Caenepeel, S., Charydczak, G., Sudarsanam, S., Hunter, T. and Manning, G. 2004, Proc. Natl. Acad. Sci., 101(32), 11707-11712.
- Min, X., Lee, B. H., Cobb, M. H. and Goldsmith, E. J. 2004, Structure, 12(7), 1303-1311.
- 37. Endicott, J. A. and Noble, M. E. 2013, Biochem. Soc. Trans., 41(4), 1008-1016.